

Application of Pyrolysis–Gas Chromatography–Mass Spectrometry for the Identification of Polymeric Materials



Peter Kusch¹, Gerd Knupp¹, Wolfgang Fink¹, Dorothee Schroeder-Obst¹, Volker Obst², and Johannes Steinhaus¹,

¹Hochschule Bonn-Rhein-Sieg, University of Applied Sciences, Department of Applied Natural Sciences, Rheinbach, Germany, ²Volker Obst, Dr. Obst Technische Werk-stoffe GmbH, Rheinbach, Germany.

The analytical pyrolysis technique hyphenated to gas chromatography–mass spectrometry (GC–MS) has extended the range of possible tools for the characterization of synthetic polymers and copolymers. Pyrolysis involves thermal fragmentation of the analytical sample at temperatures of 500–1400 °C. In the presence of an inert gas, reproducible decomposition products characteristic for the original polymer or copolymer sample are formed. The pyrolysis products are chromatographically separated using a fused-silica capillary column and are subsequently identified by interpretation of the obtained mass spectra or by using mass spectra libraries. The analytical technique eliminates the need for pretreatment by performing analyses directly on the solid or liquid polymer sample. In this article, application examples of analytical pyrolysis hyphenated to GC–MS for the identification of different polymeric materials in the plastic and automotive industry, dentistry, and occupational safety are demonstrated. For the first time, results of identification of commercial light-curing dental filling material and a car wrapping foil by pyrolysis–GC–MS are presented.

Structural analysis and the study of degradation properties are important to understand and improve performance characteristics of synthetic polymers and copolymers in many industrial applications. Traditional analytical techniques used for characterization of polymers and copolymers such as thermal analysis and Fourier transform infrared (FT–IR) spectroscopy have limitations or are not sufficiently sensitive (1). Pyrolysis techniques hyphenated to gas chromatography–mass spectrometry (GC–MS) have extended the range of possible tools for the characterization of synthetic polymers and copolymers. Under controlled conditions, at elevated temperatures (500–1400 °C) in the presence of an inert gas, reproducible decomposition products characteristic for the original polymer or copolymer sample are formed. The pyrolysis products are chromatographically separated using a fused-silica capillary column and subsequently identified by interpretation of the obtained mass spectra or by using mass spectra libraries (such as the National Institute of Standards and Technology [NIST] or Wiley). Pyrolysis methods eliminate the need for pretreatment by performing analyses directly on the solid polymer or copolymer sample (1). (Please note that this article was presented at the XVII European Conference on Analytical Chemistry, which was held in Warsaw, Poland, on 25–29 August 2013).

Most of the thermal degradation results from free radical reactions initiated by bond breaking and depends on the relative strengths of the bonds that hold the molecules together. A large molecule will break apart and rearrange in a characteristic

way (2–4). If the energy transfer to the sample is controlled by temperature, heating rate, and time, the fragmentation pattern is reproducible and characteristic for the original polymer or copolymer. Another sample of the same composition, heated at the same rate to the same temperature for the same period of time, will produce the same decomposition products. Therefore, the essential requirements of the apparatus in analytical pyrolysis are reproducibility of the final pyrolysis temperature, rapid temperature rise, and accurate temperature control. Depending on the heating mechanism, pyrolysis systems have been classified into two groups: the continuous-mode pyrolyzer (furnace pyrolyzer) and pulse-mode pyrolyzer (flash pyrolyzer, such as the heated filament, Curie-point, and laser pyrolyzer). The pyrolysis unit is directly connected to the injector port of a gas

KEY POINTS

- Pyrolysis–GC–MS is a valuable technique for the analysis and identification of synthetic polymers and copolymers.
- The technique described allows the direct analysis of very small sample amounts (5–200 µg) without the need for time-consuming sample preparation.
- Commercial light-curing dental filling material and car wrapping foil were identified using this method.

Figure 1: Schematic view of the furnace pyrolyzer used in this study.

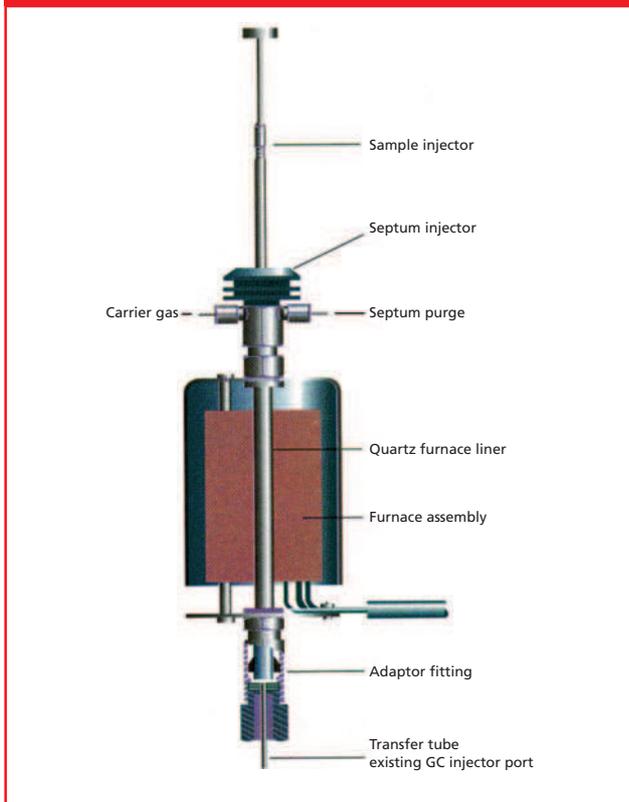
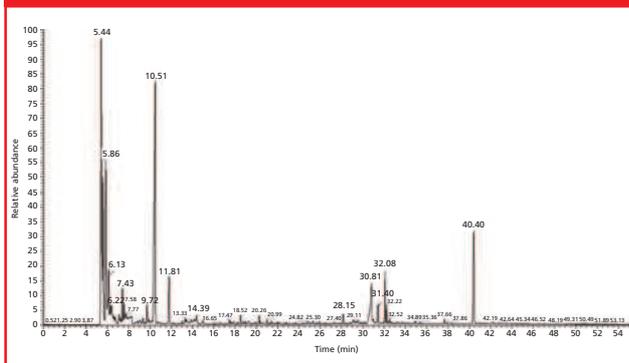


Figure 2: Pyrolysis–GC–MS chromatogram of plastic particles from industrial filter fins at 700 °C obtained with apparatus 1. Fused-silica GC capillary column: 60 m × 0.25 mm, 0.25- μ m ϕ Elite-5ms. GC conditions: programmed column temperature: 60 °C for 1 min, then 60–100 °C at 2.5 °C/min and then 100–280 °C at 10 °C/min (20 min hold at 280 °C); split–splitless injector temperature: 250 °C; split flow: 50 cm³/min; helium programmed pressure: 70 kPa for 1 min, then 70–110 kPa at 1 kPa/min (hold at 110 kPa to the end of analysis). For peak identification, see Table 1.



chromatograph. A flow of an inert carrier gas, such as helium, flushes the pyrolyzates into the fused-silica capillary column. Figure 1 shows the schematic view of the furnace pyrolyzer used in our investigation. The detection technique of the separated compounds is typically MS, but other GC detectors have also been used depending on the intentions of the analysis (1,4).

Table 1: Pyrolysis products and identified materials in plastic particles from industrial filter fins.

Retention Time t_R (min)	Pyrolysis Product at 700 °C	Matching Factor	Identified Material
5.44	Propylene	820	Polypropylene glycol
5.58	1-Butene/1,3-butadiene	840	Styrene–butadiene rubber (SBR)
5.86	Acetone	850	Polypropylene glycol
6.13	Pentadiene	885	SBR
7.43	Benzene	954	SBR
9.72	Toluene	863	SBR
10.51	Cyclopentanone	933	Poly(hexamethylene adipamide) (nylon 6-6)
11.81	2-Cyclopenten-1-one	906	Poly(hexamethylene adipamide) (nylon 6-6)
14.39	Styrene	851	SBR
28.15	4-Isopropylphenol	944	Polycarbonate or bisphenol A epoxy resin
40.40	<i>N</i> -Phenyl-1-naphthalen-amine	948	Antioxidant

The applications of analytical pyrolysis–GC–MS range from research and development of new materials, quality control, characterization and competitor product evaluation, medicine, biology and biotechnology, geology, airspace, and environmental analysis to forensic purposes or conservation and restoration of cultural heritage. These applications cover analysis and identification of polymers, copolymers, and additives in components of automobiles, tyres, packaging materials, textile fibres, coatings, half-finished products for electronics, paints or varnishes, lacquers, leather, paper or wood products, food, pharmaceuticals, surfactants, and fragrances.

Our earlier publications (1,5–12) presented the analysis and identification of degradation products of commercially available synthetic polymers and copolymers by using analytical pyrolysis hyphenated to gas chromatography with flame ionization detection (GC–FID) and GC–MS. In this work, new examples of applications of this analytical technique for the identification of different polymeric materials are demonstrated.

Experimental

Samples: Plastic particles from industrial filter fins, a car wrapping foil, unknown fibres, and commercial light-curing dental filling material were used in the investigation.

Instrumentation and Analytical Conditions: Approximately 100–200 μ g of solid sample was cut out with a scalpel and inserted without any further preparation into the bore of the pyrolysis solids-injector and then placed with the plunger on the quartz wool of the quartz tube of the furnace pyrolyzer Pyrojector II (SGE Analytical Science). Three spots on each sample were analyzed in duplicate. The pyrolyzer was operated at a constant temperature of 550, 600, 700, or 900 °C. The pressure of helium carrier gas at the inlet to the furnace was 95 kPa.

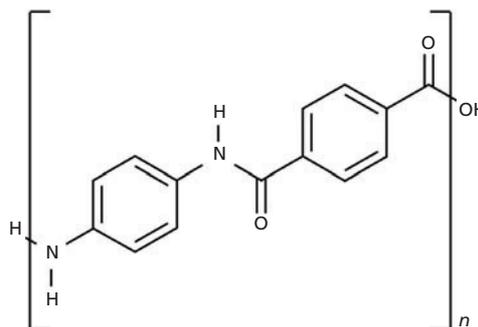
Pyrolysis–GC–MS measurements were made using two apparatus. In the first apparatus (1), the pyrolyzer was connected to a Trace 2000 gas chromatograph (ThermoQuest, CE Instruments) with a quadrupole mass spectrometer Voyager (ThermoQuest, Finnigan, MassLab Group) operated in electron ionization (EI) mode. A 60 m × 0.25 mm, 0.25- μ m Elite-5ms fused-silica GC capillary column (PerkinElmer Instruments) was used. The GC conditions were as follows: programmed

Table 2: Pyrolysis products and identified materials in car wrapping foil.

Retention time t_R (min)	Pyrolysis product at 700 °C	Matching factor	Identified material
5.49	Hydrogen chloride	945	Poly(vinyl chloride) (PVC)
5.58	Methyl chloride	800	PVC
5.64	1-Butene	938	PVC
6.01	1,3-Pentadiene	921	PVC
6.99	Tetrahydrofuran	769	Solvent
7.29	1,4-Cyclohexadiene	923	PVC
7.48	Benzene	945	PVC
8.20	Methyl methacrylate	750	Poly(methyl methacrylate) (PMMA)
9.78	Toluene	904	PVC
10.29	2-Ethyl-1-hexene	890	Bis(2-ethylhexyl) phthalate (plasticizer)
10.46	Cyclopentanone	912	Poly(hexamethylene adipamide) (nylon 6-6)
10.53	1-Octene	907	PVC
14.47	Styrene	934	PVC
17.33	2-Ethylhexanal	856	Bis(2-ethylhexyl) phthalate (plasticizer)
20.57	2-Ethyl-1-hexanol	916	Bis(2-ethylhexyl) phthalate (plasticizer)
21.07	<i>o</i> -Methylstyrene	888	PVC
21.43	Indene	870	PVC
22.99	<i>p</i> - <i>tert</i> -Butyltoluene	856	2,6-Bis-(1,1-dimethylethyl)-4-methylphenol (BHT) (antioxidant) (?)
25.93	Naphthalene	920	PVC
28.46	2-Methylnaphthalene	862	PVC
28.63	Phthalic anhydride	906	Bis(2-ethylhexyl) phthalate (plasticizer)
28.79	1-Methylnaphthalene	875	PVC
36.69	3,3-Diphenylacrylonitrile	937	Adhesive layer

($t_R = 17.33$ min), 2-ethyl-1-hexanol ($t_R = 20.57$ min), and phthalic anhydride ($t_R = 28.63$ min) (1,7). In the car wrapping material, the rest of the tetrahydrofuran solvent ($t_R = 6.99$ min) was also detected. Table 2 shows the identified ingredients of the pyrolyzed car wrapping foil.

Identification of Unknown Plastic Fibres: A sample of unknown plastic fibres was pyrolyzed at 700 °C and 900 °C, respectively, to identify its composition. Figure 4 shows the pyrolysis–GC–MS chromatogram of the sample pyrolyzed at 900 °C. Based on the decomposition products summarized in Table 3, the fibres were identified as polyaramid [poly(*p*-phenylene terephthalamide)] (Figure 5). The main identified degradation products of polyaramid at 900 °C are benzene ($t_R = 7.56$), aniline ($t_R = 11.61$ min), and benzonitrile ($t_R = 11.81$ min). Currently, polyaramid fibres have only been characterized in a few publications using thermal analysis

Figure 5: Chemical structure of poly(*p*-phenylene terephthalamide) (polyaramid).**Table 3:** Pyrolysis products of polyaramid fibres.

Peak Number	Retention Time t_R (min)	Pyrolysis Product at 900 °C	Matching Factor
1	6.52	Carbon dioxide	957
2	6.70	Acrylonitrile	896
3	7.56	Benzene	968
4	8.50	Toluene	918
5	10.20	Styrene	924
6	11.30	Isocyanatobenzene	947
7	11.61	Aniline	959
8	11.81	Benzonitrile	967
9	16.98	1,2-Benzodinitrile	959
10	17.02	1,4-Benzodiamine	905
11	19.60	Biphenyl	957
12	26.19	Acridine	931
13	26.64	1,1'-Biphenyl-4-amine	960
14	28.31	Carbazole	928
15	30.01	<i>N</i> -Phenylbenzamide	916

(thermogravimetry, derivative thermogravimetry, and differential thermal analysis), infrared spectroscopy techniques (13–15), and pyrolysis–GC–MS (2,16–19).

Polyaramid fibres are a class of heat-resistant, strong synthetic fibres. They are used in aerospace and military applications for ballistic-rated body armour, fabric, ballistic composites, and fire fighters protective clothing as well as in bicycle tyres and as an asbestos substitute.

Identification of Commercial Light-Curing Dental Filling Material:

A number of dental filling materials are presently available for tooth restorations. The four main groups of these materials, which dentists have used for about 35 years, are the conventional glass-ionomer cements, resin-based composites, resin-modified glass-ionomer cements, and polyacid-modified resinous composites (20). Light-curing glass-ionomer cements contain polyacrylic acid, chemically or photo-curing monomers (multifunctional methacrylates, like triethylene glycol dimethacrylate or 2-hydroxyethyl methacrylate), an ion-leaching glass, and additives (initiators, inhibitors, stabilizers, and others) (20). Resin-modified glass-ionomer cements are now widely used in dentistry as direct filling materials, liners, bases, luting cements, and fissure sealants (21). These materials mainly consist of polymer matrix and glass-ionomer parts. The polymer matrix is based on a monomer system and different multifunctional methacrylates with additives (21). Methacrylic monomers, such as bisphenol A glycidyl methacrylate (Bis-GMA), urethane

Figure 6: Pyrolysis–GC–MS chromatogram of commercial light-curing dental filling material at 550 °C obtained with apparatus 2. Fused-silica GC capillary column: 59 m × 0.25 mm, 0.25- μ m d_f DB-5ms. GC conditions: programmed column temperature: 60 °C for 1 min, then 60–280 °C at 7 °C/min (hold at 280 °C to the end of analysis); programmed helium pressure: 122.2 kPa for 1 min, then 122.2–212.9 kPa at 7 kPa/min (hold at 212.9 kPa to the end of analysis). For peak identification, see Table 4.

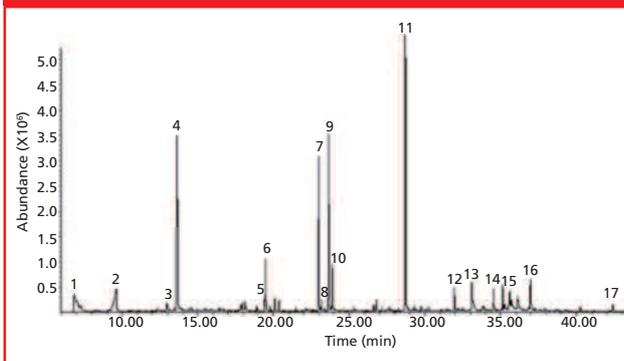


Table 4: Pyrolysis products of commercial light-curing dental filling material.

Peak Number	Retention Time t_R (min)	Pyrolysis Product at 550 °C	Matching Factor
1	6.85	Carbon dioxide	999
2	9.62	Methacrylic acid	936
3	12.96	Phenol	909
4	13.65	2-Hydroxyethyl methacrylate (HEMA)	918
5	19.40	4-Isopropenylphenol	928
6	19.48	Ethylene glycol dimethacrylate (EGDMA)	915
7	23.00	Not identified	
8	23.17	2,6-Bis-(1,1-dimethylethyl)-4-methylphenol (BHT)	923
9	23.65	Not identified	
10	23.89	Not identified	
11	28.72	Triethylene glycol dimethacrylate (TEDMA)	958
12	31.95	Drometrizol (Tinuvin-P)	938
13	33.10	4,4'-Dihydroxy-2,2-diphenylpropane (bisphenol A)	924
14	34.55	Triphenylantimony	911
15	35.25	Tetraethylene glycol dimethacrylate	791
16	36.98	Not identified	
17	42.42	Bisphenol A diglycidyl ether (BADGE)	839

dimethacrylate (UDMA), triethylene glycol dimethacrylate (TEGDMA), and 2-hydroxyethyl methacrylate (HEMA), are the main components of resin-based dental filling materials. The presence of additives such as initiators, activators, inhibitors, and plasticizers in uncured dental material mixture is necessary (21).

Figure 6 shows the total ion current pyrolysis–GC–MS chromatogram of commercial light-curing dental filling material pyrolyzed at 550 °C. The pyrolysis products identified by using mass spectra library NIST 05 are summarized in Table 4.

The carbon dioxide ($t_R = 6.85$ min) identified in pyrolyzate is formed from polyacrylic acid (2,18). The identified substances 2-hydroxyethyl methacrylate (HEMA) ($t_R = 13.65$ min), ethylene glycol dimethacrylate (EGDMA) ($t_R = 19.48$ min), and triethylene glycol dimethacrylate (TEDMA) ($t_R = 28.72$ min) are known as standard composites of dental filling materials (1). Other compounds in Table 4, such as bisphenol A ($t_R = 33.10$ min) or bisphenol A diglycidyl ether ($t_R = 42.42$ min), are probably formed by thermal degradation of bisphenol A diglycidyl mono- or dimethacrylates. The presence of the additives, such as the antioxidant 2,6-bis(1,1-dimethylethyl)-4-methylphenol (BHT) ($t_R = 23.17$ min) or the UV-absorber drometrizol ($t_R = 31.95$ min) was also confirmed. The triphenylantimony ($t_R = 34.55$ min) identified in pyrolyzate is used as catalyst in the UV-induced polymerization (1).

Conclusion

Analytical pyrolysis–GC–MS has been proven as a valuable technique for the analysis and identification of organic polymeric materials in the plastic and rubber industry. For the first time pyrolysis–GC–MS was used for the identification of commercial light-curing dental filling material and for the identification of a car wrapping foil. This technique allows the direct analysis of very small sample amounts (5–200 μ g) without the need for time-consuming sample preparation.

References

- (1) P. Kusch, in *Advanced Gas Chromatography – Progress in Agricultural, Biomedical and Industrial Applications*, M.A. Mohd, Ed. (InTech, Rijeka, Croatia, 2012), pp. 343–362.
- (2) S.C. Moldoveanu, *Analytical Pyrolysis of Synthetic Organic Polymers* (Elsevier, Amsterdam, The Netherlands, 2005).
- (3) T.P. Wampler, *Applied Pyrolysis Handbook, 2nd Ed.* (CRC Press, Boca Raton, Florida, USA, 2007).
- (4) K.L. Sobehi, M. Baron, and J. Gonzales-Rodrigues, *J. Chromatogr. A* **1186**, 51–66 (2008).
- (5) P. Kusch, *Chem. Anal. (Warsaw)* **41**, 241–252 (1996).
- (6) P. Kusch, G. Knupp, and A. Morisson, in *Horizons in Polymer Research*, R.K. Bregg, Ed. (Nova Science Publishers, New York, New York, USA, 2005), pp. 141–191.
- (7) P. Kusch, *LCGC North Am.* **31**(3), 248–254 (2013).
- (8) P. Kusch, V. Obst, D. Schroeder-Obst, W. Fink, G. Knupp, and J. Steinhaus, *Eng. Fail. Anal.* **35**, 114–124 (2013).
- (9) P. Kusch and G. Knupp, *Nachr. Chem.* **57**, 682–685 (2009).
- (10) P. Kusch, V. Obst, D. Schroeder-Obst, G. Knupp, and W. Fink, *LCGC AdS*, July/August, 5–11 (2008).
- (11) P. Kusch and G. Knupp, *LCGC AdS*, June, 28–34 (2007).
- (12) P. Kusch, W. Fink, D. Schroeder-Obst, and V. Obst, *Aluminium (Isernhagen, Germany)* **84**(4), 76–79 (2008).
- (13) S. Villar-Rodil, A. Martínez-Alonso, and J.M.D. Tascón, *J. Anal. Appl. Pyrol.* **58–59**, 105–115 (2001).
- (14) F. Suárez-García, A. Martínez-Alonso, and J.M.D. Tascón, *Carbon* **42**, 1419–1426 (2004).
- (15) S. Villar-Rodil, A. Martínez-Alonso, and J.M.D. Tascón, *J. Therm. Anal. Calorim.* **79**, 529–532 (2005).
- (16) J.R. Brown and A.J. Power, *Polym. Degrad. Stab.* **4**(5), 379–392 (1982).
- (17) H.-R. Schulten, B. Plage, H. Ohtani, and S. Tsuge, *Angew. Makromol. Chem.* **155**, 1–20 (1987).
- (18) S. Tsuge, H. Ohtani, and C. Watanabe, *Pyrolysis-GC/MS Data Book of Synthetic Polymers* (Elsevier, Amsterdam, The Netherlands, 2011).
- (19) Zs. Czégény and M. Blazsó, *J. Anal. Appl. Pyrol.* **58–59**, 95–104 (2001).
- (20) R. Rogalewicz, A. Voelkel, and I. Kownacki, *J. Environ. Monit.* **8**, 377–383 (2006).
- (21) R. Rogalewicz, K. Batko, and A. Voelkel, *J. Environ. Monit.* **8**, 750–758 (2006).

Peter Kusch, Gerd Knupp, Wolfgang Fink, Dorothee Schroeder-Obst, and Johannes Steinhaus are with Hochschule Bonn-Rhein-Sieg, University of Applied Sciences in the Department of Applied Natural Sciences, in Rheinbach, Germany.
Volker Obst is with Dr. Obst Technische Werkstoffe GmbH, in Rheinbach, Germany.